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## CHEMICAL CONSTITUENTS OF THE LEAVES OF *PLANTAGO MAJOR* L.

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### Summary

#### Chemical Constituents of the Leaves of *Plantago major* L.

From ethyl acetate and *n*-butanol extracts of the leaves of *Plantago major* L., seven compounds (1-7) were isolated. Their chemical structures were identified as 5-(hydroxymethyl)furan-2-carbaldehyde (1), vanillic acid (2), uracil (3), daucosterol (4), baicalein (5), scutellarein-7-*O*- $\beta$ -D-glucopyranoside (6) and oct-1-en-3-*O*- $\beta$ -D-xylopyranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside (7), based on spectral data and literature references. Compounds 1, 3, 6, and 7 were isolated from this plant for the first time.

**Keywords:** *Plantago major*, Scutellarein-7-*O*- $\beta$ -D-glucopyranoside, Oct-1-en-3-*O*- $\beta$ -D-xylopyranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside.

### 1. Introduction

*Plantago major* L. belongs to the Plantaginaceae family and is widely cultivated in Vietnam for its medicinal properties, including diuretic effects, and treatment for long coughs, tracheitis, dysentery, and red eye [1]. Scientific studies *in vivo* and *in vitro* have shown wound healing effects, antidiabetic, antidiarrhoeal, anti-inflammatory, antinociceptive, antioxidant and free radical scavenger, anticancer, antibacterial, and antiviral [2],[3],[4],[5],[6],[7],[8]. The up-to-date phytochemical studies on the plant have resulted in the identification of flavonoids, alkaloids, terpenoids, phenolics, iridoid glycosides, fatty acids, polysaccharides, and vitamins [8],[9],[10],[11],[12],[13],[14],[15]. In this study, we isolated and identified seven compounds from the leaves of *Plantago major*: 5-(hydroxymethyl)furan-2-carbaldehyde (1), vanillic acid (2), uracil (3), daucosterol (4), baicalein (5), scutellarein-7-*O*- $\beta$ -D-glucopyranoside (6), and oct-1-en-3-*O*- $\beta$ -D-xylopyranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside (7)

(Fig.1). Among them, compounds 1, 3, 6 and 7 were isolated from this plant for the first time.

### Materials and methods

#### 2.1. Plant material

The samples were collected at the Hanoi Research Centre for Cultivation and Processing of Medicinal Plants in May 2023. The plant was identified *Plantago major* L., Plantagineaceae by MSc. Nguyen Van Hieu and MSc. Nguyen Quynh Nga. A voucher specimen (PM52023) was deposited at the National Institute of Medicinal Materials.

#### 2.2. General experiment procedures

The NMR measurements were conducted using CD<sub>3</sub>OD, CDCl<sub>3</sub>, and DMSO-*d*<sub>6</sub> as solvents on a Bruker NMR spectrometer with a frequency of 600 MHz. Tetramethylsilane was used as an internal standard, and chemical shifts were reported in  $\delta$  (ppm). Electrospray Ionization Mass Spectrometry (ESI-MS) was performed using an Agilent 1100 series LC-MSD ion trap spectrometer. Column chromatography was performed using *silica gel* (70-230 or 230-400

mesh, Merck) and YMC (ODS-A 12 nm, S-75  $\mu\text{m}$ , Japan) as a stationary phase. Thin Layer Chromatography (TLC) was carried out on *silica gel* 60 F<sub>254</sub> plates (Merck). Spots were detected by UV radiation (245 and 365 nm) or by spraying with 10% H<sub>2</sub>SO<sub>4</sub> or FeCl<sub>3</sub> 5% reagent, followed by heating.

### 2.3. Extraction and isolation

The dried and pulverized leaves of *P. major* (5.0 kg) were extracted with 80% ethanol (v/v) for 3 hours at 65°C, and concentrated under reduced pressure to yield total extract (TMD, 548.5 g, 11%). The total extract was suspended in water and successively extracted with *n*-hexane (nH), ethyl acetate (EtOAc), and *n*-butanol. The solvent was evaporated *in vacuo* to obtain corresponding *n*-hexane (HMD, 148.1 g), ethyl acetate (EMD, 15.5 g), *n*-butanol (BMD 97.6 g), and water (WMD, 287.0 g) extracts, respectively.

The EMD and BMD extracts were combined (EBMD, 110 g) and chromatographed on a *silica gel* column, eluting with gradient solvents of EtOAc/MeOH (50/1-5/1-100%MeOH, v/v) to give 7 fractions (A-G). The A fraction (3.9 g) was separated on a *silica gel* column, eluting with gradient solvents of DCM/MeOH (15/1-5/1, v/v) to afford 2 fractions (A1 and A2) and compound **4** (17 mg). Compound **1** (8 mg) was yielded from the A1 fraction (157 mg) using a *silica gel* column with an eluent of *n*-hexane/acetone (3/1, v/v). The A2 fraction (527 mg) was continuously separated on a *silica gel* column, eluting with solvents of *n*-hexane/MeOH (1/1, v/v) to afford A2.1 precipitation and A2.2 fraction. Compound **2** (7 mg) was yielded from crystallizing and washing A2.1 precipitation (36 mg) with an eluent of DCM/MeOH (15/1, v/v). Compound **3** (6 mg) was isolated from the A2.2 fraction (43 mg) by YMC column with an eluent of MeOH/water (10/1, v/v). The B fraction (1.5 g) was separated on a *silica gel* column, eluting with solvents of *n*-hexane/acetone (2/1, v/v) to obtain three fractions (B1-B3). Compound **5** (12 mg) was yielded from the B1 fraction (76 mg) by the YMC column with an eluent of MeOH/water (4/1, v/v). The E fraction (5.7 g) was chromatographed on a *silica gel* column, eluting with gradient solvents of DCM/MeOH (10/1-5/1, v/v) to give E1 precipitation and 2 fractions (E2

and E3). Compound **6** (17 mg) was yielded from crystallizing and washing E1 precipitation (810 mg) with an eluent of DCM/acetone (2/1, v/v). Compound **7** (6 mg) was yielded from the E3 fraction (48 mg) by the YMC column with an eluent of MeOH/water (2/1, v/v).

Compound **1** (5-(hydroxymethyl)furan-2-carbaldehyde): white powder; ESI-MS *m/z*: 127.2 [M+H]<sup>+</sup>, C<sub>6</sub>H<sub>6</sub>O<sub>3</sub>. <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)  $\delta_{\text{H}}$  (ppm): 9.61 (1H, s, H-6), 7.22 (1H, d, *J* = 3.6 Hz, H-3), 6.52 (1H, d, *J* = 3.6 Hz, H-4), 4.73 (2H, s, H-7). <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)  $\delta_{\text{C}}$  (ppm): 177.6 (C-6), 160.4 (C-5), 152.5 (C-2), 124.1 (C-3), 110.0 (C-4), 57.7 (C-7).

Compound **2** (vanillic acid): white crystal. ESI-MS *m/z*: 169.2 [M+H]<sup>+</sup>. <sup>1</sup>H-NMR (600 MHz, CD<sub>3</sub>OD)  $\delta_{\text{H}}$  (ppm): 7.60 (1H, d, *J* = 1.8 Hz, H-2), 7.50 (1H, dd, *J* = 1.8, 7.8 Hz, H-6), 6.77 (1H, d, *J* = 7.8 Hz, H-5), 3.90 (3H, s, 3-OCH<sub>3</sub>). <sup>13</sup>C-NMR (150 MHz, CD<sub>3</sub>OD)  $\delta_{\text{C}}$  (ppm): 175.4 (C-7), 150.0 (C-3), 148.1 (C-4), 128.3 (C-1), 124.2 (C-6), 115.2 (C-5), 114.1 (C-2), 56.3 (3-OCH<sub>3</sub>).

Compound **3** (uracil): white powder. Mp: 335°C. ESI-MS *m/z*: 113.2 [M+H]<sup>+</sup>. <sup>1</sup>H-NMR (600 MHz, CD<sub>3</sub>OD)  $\delta_{\text{H}}$  (ppm): 7.41 (1H, d, *J* = 7.2 Hz, H-6), 5.63 (1H, d, *J* = 7.2 Hz, H-5). <sup>13</sup>C-NMR (150 MHz, CD<sub>3</sub>OD)  $\delta_{\text{C}}$  (ppm): 167.3 (C-4), 153.5 (C-2), 143.5 (C-6), 101.7 (C-5).

Compound **4** (daucosterol): white powder; <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub> & CD<sub>3</sub>OD)  $\delta_{\text{H}}$  (ppm): 5.37 (1H, m, H-6), 3.60 (1H, m, H-3), 1.03 (3H, s, H-19), 0.94 (3H, d, *J* = 6.6 Hz, H-21), 0.86 (3H, t, *J* = 7.8 Hz, H-29), 0.85 (3H, d, *J* = 6.6 Hz, H-27), 0.83 (3H, d, *J* = 6.6 Hz, H-26), 0.71 (3H, s, H-18). Glc: 4.40 (1H, d, *J* = 7.8 Hz, H-1'), 3.86 (1H, dd, *J* = 12.0, 3.0 Hz, H-6'), 3.76 (1H, dd, *J* = 12.0, 5.4 Hz, H-6'), 3.38 (2H, m, H-3', H-4'), 3.27 (1H, m, H-5'), 3.20 (1H, m, H-2').

Compound **5** (baicalein): yellow powder. ESI-MS *m/z*: 271.1 [M+H]<sup>+</sup>. <sup>1</sup>H-NMR (600 MHz, acetone-*d*<sub>6</sub>)  $\delta_{\text{H}}$  (ppm): 12.73 (1H, s, 5-OH), 8.05 (2H, dd, *J* = 1.5, 8.0 Hz, H-2', H-6'), 7.56 - 7.61 (3H, m, H-3', H-4', H-5'), 6.76 (1H, s, H-3), 6.69 (1H, s, H-8). <sup>13</sup>C-NMR (150 MHz, acetone-*d*<sub>6</sub>)  $\delta_{\text{C}}$  (ppm): 183.4 (C-4), 164.6 (C-2), 153.8 (C-7), 151.5 (C-9), 148.0 (C-5), 132.5 (C-1', C-6), 132.4 (C-4'), 129.9 (C-3', C-5'), 127.1 (C-2', C-6'), 105.4 (C-3), 105.7 (C-10), 94.8 (C-8).

Compound **6** (scutellarein-7-*O*- $\beta$ -D-glucopyranoside): white powder; ESI-MS *m/z*:

447.5 [M-H]<sup>+</sup>. <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)  $\delta_H$  (ppm): 7.93 (2H, d, *J* = 9.0 Hz, H-2', H-6'), 7.00 (1H, s, H-8), 6.94 (2H, d, *J* = 9.0 Hz, H-3' & H-5'), 6.81 (1H, s, H-3), 5.00 (1H, d, *J* = 7.2 Hz, H-1''), 3.75 (1H, dd, *J* = 4.2, 11.4 Hz, H-6a''), 3.51 (1H, dd, *J* = 5.4, 11.4 Hz, H-6b''). <sup>13</sup>C-NMR (150 MHz, DMSO-*d*<sub>6</sub>)  $\delta_C$  (ppm): 182.3 (C-4), 164.0 (C-2), 161.2 (C-4'), 151.3 (C-9), 149.0 (C-7), 146.5 (C-5), 130.4 (C-6), 128.4 (C-2', C-6'), 121.2 (C-1'), 115.9 (C-3', C-5'), 105.8 (C-10), 102.4 (C-3), 101.0 (C-1''), 94.1 (C-8), 77.3 (C-5''), 75.8 (C-3''), 73.1 (C-2''), 69.7 (C-4''), 60.6 (C-6'').

Compound 7 (oct-1-en-3-*O*- $\beta$ -D-xylopyranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside): ESI-MS *m/z*: 421.5 [M-H]<sup>+</sup>. <sup>1</sup>H-NMR (600 MHz,

CD<sub>3</sub>OD)  $\delta_H$  (ppm): 5.89 (1H, ddd, *J* = 6.6, 10.8, 17.4 Hz, H-2), 5.24 (1H, ddd, *J* = 1.2, 1.7, 17.4 Hz, H-1), 5.13 (1H, ddd, *J* = 1.2, 1.7, 11.0 Hz, H-1), 4.36 (1H, d, *J* = 7.8 Hz, H-1''), 4.34 (1H, d, *J* = 7.8 Hz, H-1'), 4.16 (1H, dd, *J* = 7.2, 13.2 Hz, H-3), 4.04 (1H, dd, *J* = 2.4, 12.0 Hz, H-6'a), 3.87 (1H, dd, *J* = 5.4, 11.4 Hz, H-5a''), 3.74 (1H, dd, *J* = 5.4, 11.4 Hz, H-6'b), 3.22 (1H, overlap, H-5b''), 1.52 - 1.58 and 1.67 - 1.73 (2H, m, H-4), 1.31 - 1.39 (6H, m, H-5, H-6, H-7), 0.92 (3H, t, *J* = 6.9 Hz, H-8). <sup>13</sup>C-NMR (150 MHz, CD<sub>3</sub>OD)  $\delta_C$  (ppm): 140.9 (C-2), 116.2 (C-1), 105.3 (C-1'), 103.3 (C-1'), 82.7 (C-3), 78.1 (C-3'), 77.6 (C-5'), 77.0 (C-3''), 75.3 (C-2'), 74.9 (C-2''), 71.5 (C-4''), 71.2 (C-4'), 69.5 (C-6'), 66.8 (C-5''), 35.7 (C-4), 33.0 (C-6), 25.7 (C-5), 23.6 (C-7), 14.4 (C-8).

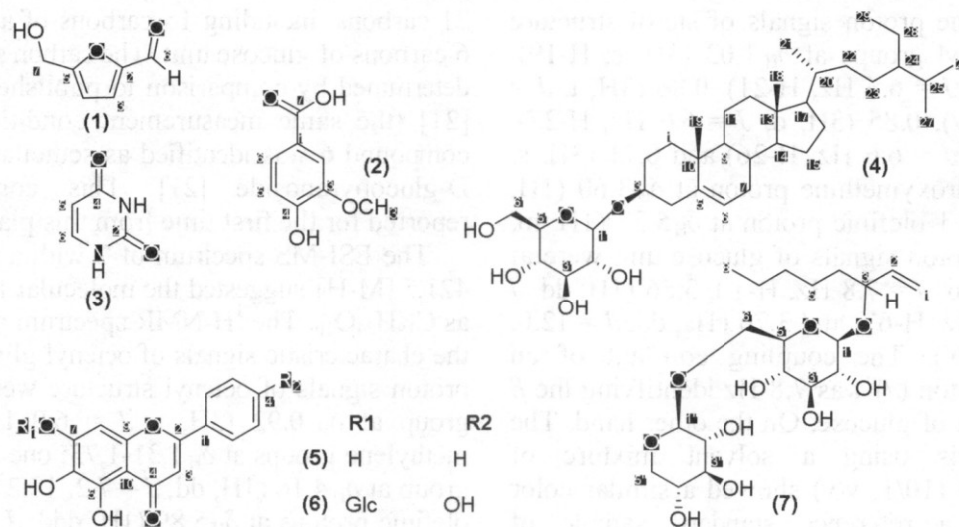


Fig. 1. Chemical structures of compounds (1-7)

### 3. Results and discussion

Compound 1 was isolated as a white powder and has a molecular formula of C<sub>6</sub>H<sub>6</sub>O<sub>3</sub> based on the ESI-MS spectrum with a peak at *m/z* 127.2 [M+H]<sup>+</sup>. The <sup>1</sup>H-NMR spectrum of 1 showed an aldehyde group at  $\delta_H$  9.61 (1H, s, H-6); two methineprotons at  $\delta_H$  7.22 (1H, d, *J* = 3.6 Hz, H-3) and 6.52 (1H, d, *J* = 3.6 Hz, H-4); and two hydroxymethylene protons  $\delta_H$  4.73 (2H, s, H-7). The <sup>13</sup>C-NMR spectrum exhibited six carbons: an aldehyde carbon at  $\delta_C$  177.6 (C-6); two methine carbons at  $\delta_C$  124.1 (C-3) and 110.0 (C-4); one hydroxymethylene carbon  $\delta_C$  57.7 (C-7); and two oxygenated carbons at  $\delta_C$  160.4 (C-5) and 152.5 (C-2). Based on these spectroscopic data and by comparison to published literature [16], compound 1 was identified as 5-(hydroxymethyl)furan-2-carbaldehyde. This compound is reported for the first time from *P. major*.

Compound 2 was isolated as a white crystal. Its molecular formula was determined to be C<sub>8</sub>H<sub>8</sub>O<sub>4</sub> based on the ESI-MS spectrum with a peak at *m/z* 169.2 [M+H]<sup>+</sup>. The <sup>1</sup>H-NMR spectrum of 2 showed the presence of 3 proton signals of aromatic ring at  $\delta_H$  7.60 (1H, d, *J* = 1.8 Hz, H-2), 7.50 (1H, dd, *J* = 1.8, 7.8 Hz, H-6) and 6.77 (1H, d, *J* = 7.8 Hz, H-5); and one methoxy group at  $\delta_H$  3.90 (3H, s, 3-OCH<sub>3</sub>). The <sup>13</sup>C-NMR spectrum displayed 8 carbons, including one carboxyl carbon at  $\delta_C$  175.4 (C-7); two oxygenated carbons at  $\delta_C$  150.0 (C-3) and 148.1 (C-4); one quaternary carbon at  $\delta_C$  128.3 (C-1); three methine carbons at  $\delta_C$  124.2 (C-6), 115.2 (C-5) and 114.1 (C-2); and one methoxy carbon at  $\delta_C$  56.3 (3-OCH<sub>3</sub>). Compound 2 was identified as vanillic acid based on the above evidence and by comparison to published literature [17].



Compound **3** was isolated as a white powder. The ESI-MS spectrum with a peak at  $m/z$  113.2  $[M+H]^+$  suggested the molecular formula of **3** as  $C_4H_4N_2O_2$ . The  $^1H$ -NMR spectrum of **3** showed the presence of two olefinic proton signals at  $\delta_H$  7.41 (1H, d,  $J = 7.2$  Hz, H-6) and 5.63 (1H, d,  $J = 7.2$  Hz, H-5). The  $^{13}C$ -NMR spectrum displayed 4 carbons, including 2 signals of C=O carbons at  $\delta_C$  167.3 (C-4) and 153.5 (C-2); 2 olefinic carbons at  $\delta_C$  143.5 (C-6) and 101.7 (C-5). Based on the above evidence and by comparison to published literature [18], compound **3** was identified as uracil. This compound is isolated from *P. major* for the first time.

Compound **4** was obtained as a yellow amorphous powder. The  $^1H$ -NMR spectrum of **4** showed the characteristic signals of sterol glucoside. The proton signals of sterol structure were 6 methyl groups at  $\delta_H$  1.03 (3H, s, H-19), 0.94 (3H, d,  $J = 6.6$  Hz, H-21), 0.86 (3H, t,  $J = 7.8$  Hz, H-29), 0.85 (3H, d,  $J = 6.6$  Hz, H-27), 0.83 (3H, d,  $J = 6.6$  Hz, H-26) and 0.71 (3H, s, H-18); 1 hydroxymethine proton at  $\delta_H$  3.60 (1H, m, H-3); and 1 olefinic proton at  $\delta_H$  5.37 (1H, m, H-6). The proton signals of glucose unit were at  $\delta_H$  4.40 (1H, d,  $J = 7.8$  Hz, H-1'), 3.86 (1H, dd,  $J = 12.0, 3.0$  Hz, H-6'), and 3.76 (1H, dd,  $J = 12.0, 5.4$  Hz, H-6'). The coupling constant of an anomeric proton ( $J$ ) was 7.8 Hz identifying the  $\beta$  configuration of glucose. On the other hand, The TLC analysis using a solvent mixture of DCM/MeOH (10/1, v/v) showed a similar color and  $R_f$  to a reference standard sample of daucosterol, which further confirmed its identity. Based on comparison to published literature [19] and the TLC with daucosterol, compound **4** was identified as daucosterol.

Compound **5** was isolated as a yellow powder. The ESI-MS spectrum with a peak at  $m/z$  271.1  $[M+H]^+$  suggested the molecular formula of **5** as  $C_{15}H_{10}O_5$ . The  $^1H$ -NMR spectrum of **5** showed the presence of 7 proton signals of aromatic rings, of which one signal at  $\delta_H$  6.69 (1H, s, H-8) belongs to A ring; one signal at  $\delta_H$  6.76 (1H, s, H-3) belongs to C ring; and 5 signals at  $\delta_H$  8.05 (2H, dd,  $J = 1.5, 8.0$  Hz, H-2', H-6') and 7.56 - 7.61 (3H, m, H-3', H-4', H-5') belong to B ring. The  $^{13}C$ -NMR spectrum of **5** displayed fifteen carbon signals including one keton carbon at  $\delta_C$  183.4 (C-4); seven non-hydrogenated carbons at  $\delta_C$  164.6 (C-2), 153.8 (C-7), 151.5 (C-9), 148.0 (C-5), 132.5 (C-1', C-6) and 105.7 (C-10); seven methine carbons at  $\delta_C$  132.4 (C-4'), 129.9 (C-3', C-5'), 127.1 (C-2', C-6'), 105.4 (C-3) and 94.8

(C-8). Based on these spectroscopic data and by comparison to published literature [20], compound **5** was identified as baicalein.

Compound **6** was isolated as a white powder. The ESI-MS spectrum with a peak at  $m/z$  447.5  $[M-H]^-$  suggested the molecular formula of **6** as  $C_{21}H_{20}O_{11}$ . The  $^1H$ -NMR spectrum of **6** was similar to that of **5**, except for additional proton signals belonging to one sugar unit, with an anomeric proton at  $\delta_H$  5.00 (1H, d,  $J = 7.2$  Hz, H-1''); a hydroxy methylene group at  $\delta_H$  3.75 (1H, dd,  $J = 4.2, 11.4$  Hz, H-6a'') and 3.51 (1H, dd,  $J = 5.4, 11.4$  Hz, H-6b''); and other proton signals of hydroxymethines of sugar unit at  $\delta_H$  3.26 - 4.67. The coupling constant of an anomeric proton ( $J$ ) was 7.2 Hz, identifying the  $\beta$  configuration of glucose. The  $^{13}C$ -NMR spectrum of **6** exhibited 21 carbons, including 15 carbons of a flavon and 6 carbons of glucose unit. The carbon signals were determined by comparison to published literature [21] (the same measurement conditions). Thus, compound **6** was identified as scutellarein-7-O- $\beta$ -D-glucopyranoside [21]. This compound is reported for the first time from this plant.

The ESI-MS spectrum of **7** with a peak at  $m/z$  421.5  $[M-H]^-$  suggested the molecular formula of **7** as  $C_{19}H_{34}O_{10}$ . The  $^1H$ -NMR spectrum of **7** showed the characteristic signals of octenyl glycoside. The proton signals of octenyl structure were 1 methyl group at  $\delta_H$  0.92 (3H, t,  $J = 6.9$  Hz, H-8); 4 methylene groups at  $\delta_H$  1.31-1.73; one oxymethine group at  $\delta_H$  4.16 (1H, dd,  $J = 7.2, 13.2$  Hz, H-3); 3 olefinic protons at  $\delta_H$  5.89 (1H, ddd,  $J = 6.6, 10.8, 17.4$  Hz, H-2), 5.24 (1H, ddd,  $J = 1.2, 1.7, 17.4$  Hz, H-1) and 5.13 (1H, ddd,  $J = 1.2, 1.7, 11.0$  Hz, H-1). The proton signals of two sugar units were 2 anomeric protons at  $\delta_H$  4.36 (1H, d,  $J = 7.8$  Hz, H-1'') and 4.34 (1H, d,  $J = 7.8$  Hz, H-1'); 2 oxygenated methylene groups at  $\delta_H$  4.04 (1H, dd,  $J = 2.4, 12.0$  Hz, H-6'a), 3.74 (1H, dd,  $J = 5.4, 11.4$ , H-6'b) and 3.87 (1H, dd,  $J = 5.4, 11.4$  Hz, H-5a''), 3.22 (1H, overlap, H-5b''); and other proton signals of hydroxymethines of two sugar units at  $\delta_H$  3.19 - 4.37. The coupling constant of two anomeric protons ( $J$ ) was 7.8 Hz, identifying the  $\beta$  configuration of two sugar units. The HMBC correlations of the anomeric proton H-1' ( $\delta_H$  4.04) with C-3 ( $\delta_C$  82.7) confirmed the attachment of the saccharide chain at C-3 of octenyl, and the anomeric proton H-1'' ( $\delta_H$  4.36) with C-6' ( $\delta_C$  69.5) verified sugar moieties was  $\beta$ -D-xylopyranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside; H-3 ( $\delta_H$  4.16) with C-2 ( $\delta_C$  140.9), C-1 ( $\delta_C$  116.2), and C-4 ( $\delta_C$  35.7) confirmed oct-1-en-3-ol (Fig. 2). Thus, compound

7 was identified as oct-1-en-3-*O*- $\beta$ -D-xylopyranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside [22]. This compound is isolated from this plant for the first time.

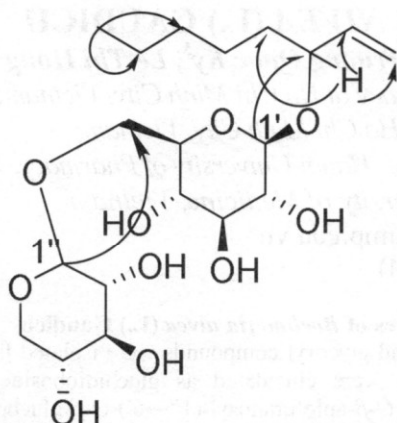


Fig. 2. The HMBC correlations of compound 7

#### 4. Conclusion

Seven compounds were obtained from the EtOAc and *n*-butanol extracts of the leaves of *P. major* including: 5-(hydroxymethyl)furan-2-carbaldehyde (1), vanillic acid (2), uracil (3), daucosterol (4), baicalein (5), scutellarein-7-*O*- $\beta$ -D-glucopyranoside (6), and oct-1-en-3-*O*- $\beta$ -D-xylopyranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside (7). The structures of these compounds were established by spectral data and literature references. Among them, compounds 1, 3, 6, and 7 were isolated from this plant for the first time.

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